



RESEARCH ARTICLE

Open Access

The effect of oxygen tension on calcium homeostasis in bovine articular chondrocytes

Rachel White and John S Gibson*

Abstract

Background: Articular chondrocytes normally experience a lower O₂ tension compared to that seen by many other tissues. This level may fall further in joint disease. Ionic homeostasis is essential for chondrocyte function but, at least in the case of H⁺ ions, it is sensitive to changes in O₂ levels. Ca²⁺ homeostasis is also critical but the effect of changes in O₂ tension has not been investigated on this parameter. Here we define the effect of hypoxia on Ca²⁺ homeostasis in bovine articular chondrocytes.

Methods: Chondrocytes from articular cartilage slices were isolated enzymatically using collagenase. Cytoplasmic Ca²⁺ levels ([Ca²⁺]_i) were followed fluorimetrically using Fura-2 to determine the effect of changes in O₂ tension. The effects of ion substitution (replacing extracellular Na⁺ with NMDG⁺ and chelating Ca²⁺ with EGTA) were tested. Levels of reactive oxygen species (ROS) and the mitochondrial membrane potential were measured and correlated with [Ca²⁺]_i.

Results: A reduction in O₂ tension from 20% to 1% for 16-18 h caused [Ca²⁺]_i to approximately double, reaching 105 ± 23 nM (p < 0.001). Ion substitutions indicated that Na⁺/Ca²⁺ exchange activity was not inhibited at low O₂ levels. At 1% O₂, ROS levels fell and mitochondria depolarised. Restoring ROS levels (with an oxidant H₂O₂, a non-specific ROS generator Co²⁺ or the mitochondrial complex II inhibitor antimycin A) concomitantly reduced [Ca²⁺]_i.

Conclusions: O₂ tension exerts a significant effect on [Ca²⁺]_i. The proposed mechanism involves ROS from mitochondria. Findings emphasise the importance of using realistic O₂ tensions when studying the physiology and pathology of articular cartilage and the potential interactions between O₂, ROS and Ca²⁺.

Background

Due to the avascularity of its matrix, articular cartilage is hypoxic compared to other tissue types [1]. O₂ tension is uncertain, but most cells probably experience 5-7% O₂ [2]. Perhaps as a consequence, articular chondrocytes have few mitochondria and metabolism is largely anaerobic. Notwithstanding, chondrocytes consume O₂ and are adversely affected if maintained in an anoxic environment [3,4]. Lowered O₂ levels can occur *in vivo* in various disease conditions [2].

It is becoming increasingly evident that O₂ tension is a critical parameter in modulating chondrocyte function [5]. At low O₂ tension, glycolysis is inhibited, glucose

uptake is reduced, and ATP and lactic acid production fall, the apparently paradoxical "negative Pasteur effect" [3]. Other responses include changes in production of growth factors, proinflammatory mediators and matrix components [5]. In other tissues, change in O₂ tension is an important signal leading to modulation of ionic permeability and alteration of ionic homeostasis, thereby impacting upon cell function [6]. Similarly, pH homeostasis in articular chondrocytes is perturbed by alteration in O₂ levels [7,8]. When O₂ is reduced from 20% to 1%, the main H⁺ efflux pathway, the Na⁺/H⁺ exchanger [9], is inhibited leading to acidification of the cells. A reduction in reactive oxygen species (ROS) acting, via alterations in protein phosphorylation, appears to constitute the link between hypoxia and reduction in NHE activity [7].

Intracellular Ca²⁺ levels are also critical [10]. Changes in Ca²⁺ will affect matrix synthesis, as well as other func-

* Correspondence: jsg1001@cam.ac.uk

¹ Department of Veterinary Medicine, University of Cambridge, Madingley Road, Cambridge, CB3 0ES, UK

Full list of author information is available at the end of the article

tions. Low O₂ tension has been shown previously to cause a rise in Ca²⁺ in cultured embryonal chick chondrocytes, acting to slow ageing processes [11]. An interaction between O₂ and Ca²⁺ is therefore anticipated in articular chondrocytes but has not been described hitherto. Our overall aim therefore was to elucidate whether Ca²⁺ levels are sensitive to O₂. Because reduction in O₂ tension from 20% to 1% has been shown to have important effects on pH homeostasis, we concentrated on these values for this study. Cytoplasmic Ca²⁺ levels, ROS and the mitochondrial membrane pd were measured fluorimetrically. Results show that Ca²⁺ levels are increased during hypoxia, with a transduction path involving mitochondrial depolarization and ROS.

Methods

Chondrocytes

Bovine feet from animals aged between 18 and 36 months were obtained following abattoir slaughter. Full depth hyaline cartilage shavings from the proximal metacarpophalangeal joint were taken at ambient O₂ tension, then placed in DMEM containing penicillin (100 IU.ml⁻¹), streptomycin (0.1 µg.ml⁻¹) and fungizone (2.5 µg.ml⁻¹) and incubated at 37°C, 5% CO₂ for 16-18 h at 20% or 1% O₂ whilst matrix was digested with 0.1% (w/v) collagenase type I. Isolated chondrocytes were resuspended in saline (at the required O₂ tension) at a final dilution of 10⁶ cells.ml⁻¹. Cell viability was determined by the Trypan Blue exclusion test, at >95%. See [12] for further details.

Solutions and chemicals

Standard saline comprised (in mM): NaCl (145), KCl (5), CaCl₂ (2), MgSO₄ (1), D⁺ glucose (10) and 4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid (HEPES, 10), pH 7.40 at 37°C. To investigate Ca²⁺-free conditions, CaCl₂ was omitted and the Ca²⁺ chelator EGTA (1 mM) added; for Na⁺-free saline, NMDG⁺ replaced Na⁺ - cells were prepared in standard saline and only exposed to these solutions for a few minutes. Stock solutions of digitonin, antimycin A and the fluorophores Fura-2, DCF-DA and JC-1 were dissolved in DMSO; CoCl₂ and H₂O₂ were dissolved in water. Fluorophores were obtained from Calbiochem (Fura-2-AM) or Molecular Probes, Invitrogen, UK; other chemicals from Sigma-Aldrich, UK.

Maintenance of O₂ tension

During longer term incubations (>3 hours), cells were maintained at the correct O₂ tension in a variable O₂/CO₂ incubator (Galaxy R, RS Biotech, Irvine, UK). For shorter term incubations, cells were placed in Eschweiler tonometers (Kiel, Germany) and flushed with appropriate gas mixtures using a Wösthoff gas mixing pump (Bochum,

Germany). Similarly, solutions were pre-equilibrated to the required O₂ tension in Eschweiler tonometers before being applied to cells.

Measurement of Ca²⁺

Cytoplasmic Ca²⁺ levels ([Ca²⁺]_i) were measured using Fura-2 (see [12]). Cells were loaded with 5 µM fura-2-AM for 30 min at room temperature followed by 15 min at 37°C. Fluorescence was measured in a thermostatically regulated fluorimeter (F-2000 Fluorescence Spectrophotometer, Hitachi). Fura-2 was alternately excited at 340 nm and 380 nm, with emission intensity was measured at 510 nm. In most cases, the 340:380 nm fluorescence ratio (R) was converted to Ca²⁺ values, as described previously [12]. When reagents were added to alter ROS levels, however, Ca²⁺ levels are presented as raw R values. In these cases, exact [Ca²⁺]_i could not be calculated because, after digitonin treatment, on exposure to the high concentrations of the reagents found extracellularly, Fura-2 was partially quenched.

Measurement of reactive oxygen species (ROS)

Chondrocytes were loaded with DCF-DA (10 µM) at 37°C for 45 min [7]. In the presence of ROS, DCF is converted to dichlorofluorescein, resulting in a change in fluorescence. DCF was excited at 488 nm and emission intensity measured at 530 nm.

Measurement of the mitochondrial pd

Chondrocytes were loaded with 5 µM JC-1 for 20 min at 37°C [8]. JC-1 was then excited at 490 nm and the emission intensity monitored at 525 nm (green) and 590 nm (red). The dye is sequestered inside mitochondria at negative pds. Membrane depolarization is indicated by a shift in the emission fluorescence from red to green, as dye is released into the cytosol and the formation of red fluorescent J-aggregates causing a fall in the red/green fluorescence intensity ratio.

Statistics

Student's paired or Independent t-test were used to determine statistical significance (p < 0.05) between results. Data are given as means ± S.E.M. for n replicates, where each replicate indicates a separate individual animal.

Results

Effect of hypoxia on Ca²⁺ homeostasis

Previously published reports on the effects of hypoxia on pH homeostasis in equine articular chondrocytes demonstrated effects within 3 hours when O₂ was reduced from 20% to 1% [7]. Evidence for a similar effect was therefore tested on Ca²⁺ levels. Bovine articular chondrocytes were isolated at 20% O₂ and the effect of maintain-

ing O₂ at this level was then compared with that of reducing it to 1% O₂. At 3 hours, [Ca²⁺]_i was 60 ± 10 nM at 20% O₂ compared with 62 ± 10 nM at 1% O₂ (means ± S.E.M., n = 12; N.S. values at 1% cf 20%). At both O₂ tensions, therefore, steady state cytoplasmic Ca levels ([Ca²⁺]_i) remained steady at about 60 nM. We went on to study the effects of longer term hypoxia. Chondrocytes were both digested from their matrix and then maintained for 16-18 hours at either 20% or 1% O₂ levels before measuring steady state Ca²⁺ levels at the same O₂ tension. At hypoxic levels, 1% O₂, a significant elevation in steady state [Ca²⁺]_i was observed (Figures 1 and 2), with levels approximately doubling from 55 ± 4 nM at 20% O₂ to 105 ± 23 nM at 1% (n = 12; p < 0.001). Thus, like pH, steady state Ca²⁺ levels in articular chondrocytes are sensitive to changes in O₂ albeit with a slower time course.

Hypoxia, Ca²⁺ and ion substitutions

Ion substitution experiments were carried out to determine the source of the extra Ca²⁺. Chondrocytes were again isolated, and then maintained for 16-18 hours, at either 20% or 1% O₂ in standard Ca²⁺- and Na⁺- containing saline. Ca²⁺ levels were then measured in this standard saline and also following transfer to Ca²⁺-free or Na⁺-free saline (Figures 1 and 2). In Ca²⁺-free conditions (Figure 1), Ca²⁺ was decreased at both 20% and 1% O₂. Notwithstanding, [Ca²⁺]_i remained higher at 1% O₂ compared to 20% O₂. In Na⁺-free saline, [Ca²⁺]_i was elevated at both O₂ tensions (Figure 2), but again remained higher at 1% O₂ compared to 20% O₂. In fact, the difference in

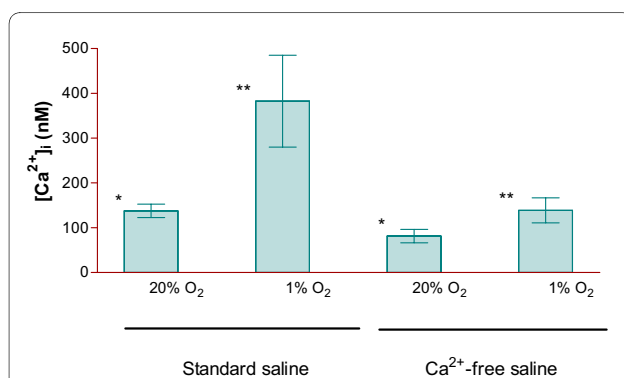


Figure 1 Effect of hypoxia and extracellular Ca²⁺ on cytoplasmic Ca²⁺ levels in bovine articular chondrocytes. Chondrocytes were isolated with collagenase at either 20% or 1% O₂ and maintained at these O₂ tensions throughout (16-18 hours). Cytoplasmic Ca²⁺ levels ([Ca²⁺]_i) were then measured with Fura-2 in the presence (2 mM Ca²⁺) or absence (Ca²⁺-free plus 1 mM EGTA) extracellular Ca²⁺. Histograms represent means ± S.E.M., n = 9. * p < 0.02 ** p < 0.006.

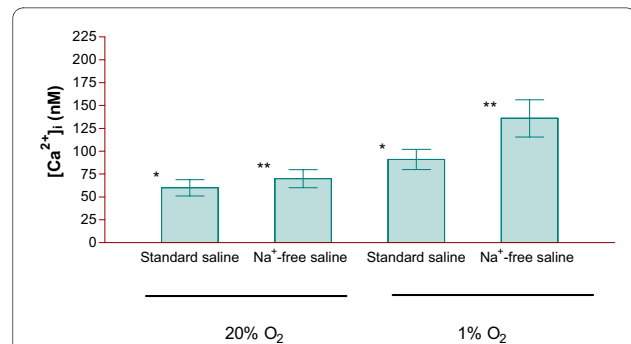


Figure 2 Effect of hypoxia and extracellular Na⁺ on cytoplasmic Ca²⁺ levels in bovine articular chondrocytes. Methods as legend to Figure 1, except that during measurement of [Ca²⁺]_i, chondrocytes were suspended in the presence (145 mM) or absence (Na⁺ replaced with NMDG⁺) of extracellular Na⁺. Histograms represent means ± S.E.M., n = 9. * p < 0.05 ** p < 0.02.

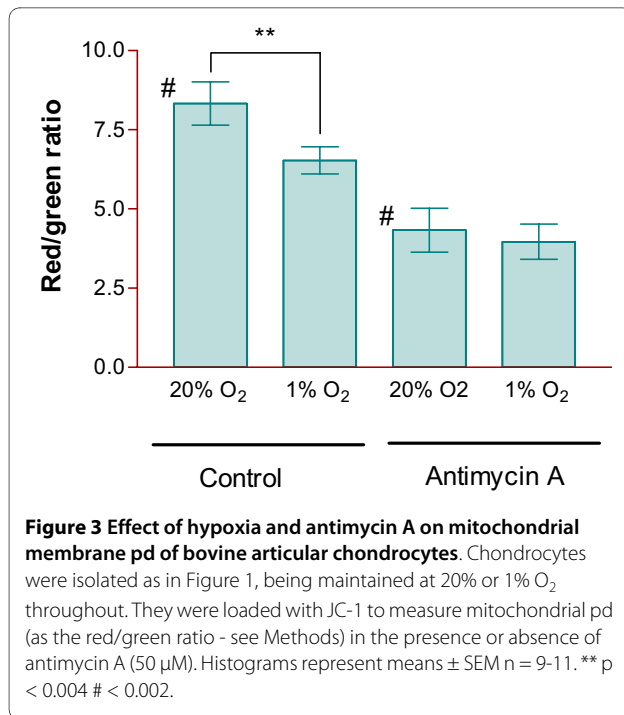
Ca²⁺ comparing cells maintained at 20% and 1% O₂ was greater in Na⁺-free conditions.

Interaction of reactive oxygen species and Ca²⁺ homeostasis

Levels of reactive oxygen species (ROS) in equine articular chondrocytes decrease when O₂ tension is reduced from 20% to 1% [7]. This finding was confirmed in the present work for bovine chondrocytes held at different O₂ levels for 16-18 hours. ROS levels at 1% fell to 60 ± 6% (mean ± S.E.M., n = 3) of the value at 20% O₂. Three different protocols were carried out to elevate ROS levels: treatment with the oxidant H₂O₂ (100 μM), the non-specific ROS generator Co²⁺ (100 μM) or the mitochondrial complex III inhibitor antimycin A (50 μM). In each case, ROS levels recorded in treated cells incubated at 1% O₂ were restored to those observed at 20%, (eg for Co²⁺ levels reached 96 ± 8% values at 20%, N.S.). Using Fura-2 340 nm:380 nm emission ratio (R) as a measure of [Ca²⁺]_i, in cells incubated at 1% but treated to raise ROS levels, it was found that R decreased by a similar amount, reaching values similar to those observed at 20%. For example, R at 1% following addition of H₂O₂ fell from 1.41 ± 0.001 to 1.06 ± 0.001 (n = 15). For all three protocols, therefore, at 1% O₂ when ROS levels were restored, so was [Ca²⁺]_i.

Hypoxia and mitochondria

The effect of changes in O₂ and treatment with antimycin A on mitochondrial pd was then investigated. Chondrocytes were isolated at 20% O₂ and then incubated at either 20% O₂ or 1% O₂ for 16-18 hours prior to loading with JC-1. They were also treated with antimycin A (50 μM) at both O₂ tensions (Figure 3). It can be seen that the red/green ratio was reduced at 1% O₂ indicative of mitochon-



drial depolarization. Antimycin A, a complex III inhibitor, also caused mitochondrial depolarization at 20% O₂ but not in cells held at 1% O₂.

Discussion

The effect of O₂ tension on steady state Ca²⁺

The present findings are the first to demonstrate an effect of changes in O₂ tension on Ca²⁺ homeostasis in articular chondrocytes. We show here that Ca²⁺ homeostasis is maintained in response to shorter term (3 hours) reduction in O₂ tension from 20% to 1%. Longer exposure to 1% O₂, however, caused significant elevation in [Ca²⁺]_i with levels approximately doubling, sufficient to perturb cell function. These effects were associated with both mitochondrial depolarization and a fall in levels of reactive oxygen species (ROS).

Source of Ca²⁺

Rise in [Ca²⁺]_i can occur through increased entry or decreased removal across the plasma membrane or from intracellular stores. It is not easy to distinguish unequivocally between these possibilities. Despite a decrease in [Ca²⁺]_i in Ca²⁺-free saline, however, hypoxic chondrocytes still showed higher Ca²⁺ compared to those at 20% O₂. Thus even if increased influx across the plasma membrane was involved, other mechanisms were still able to elevate Ca²⁺ during hypoxia. Substitution of extracellular Na⁺ increased [Ca²⁺]_i and exacerbated the difference at the two O₂ tensions. This finding is consistent with ele-

vated activity of NCE at low O₂, perhaps in an attempt to reduce Ca²⁺ to levels found at 20% O₂. Since NCE activity requires a functional ATP-driven Na⁺/K⁺ pump, it is unlikely that ATP was limiting (as shown previously [7]). In addition, because inhibition of the mitochondrial electron transport chain with antimycin A reduces [Ca²⁺]_i, any Ca²⁺ release from mitochondrial stores following their hypoxia-induced depolarization, would likely to be insufficient on its own to raise [Ca²⁺]_i. In this context, it is important to note that mitochondria in articular chondrocytes occupy a relatively small volume (1-2% cytoplasm) [13] compared to that seen in other tissues (typically 15-20%, eg liver). There is also some reduction in mitochondrial volume with depth and age [14,15]. They may also lack a functional electron transport chain [16], relying on glycolysis for metabolic energy [3]. Taken together, these findings are consistent with hypoxic release of Ca²⁺ into the cytoplasm from intracellular non-mitochondrial stores, probably endoplasmic reticulum.

Oxygen and chondrocyte function

As noted above, it is unlikely that articular chondrocytes require O₂ for energy, at least directly. Nevertheless, O₂ tension is a critical parameter in modulating chondrocyte function. Changes in O₂ level affect ATP production [3], growth factors [17], proinflammatory mediators [18] and matrix components [19]. Dedifferentiation of chondrocytes occurs when they are maintained at abnormally high O₂. This includes restoration of the ability to carry out oxidative phosphorylation [20]. Standard chondrocyte markers, such as collagen type II and aggrecan, are affected [19]. In effect, low O₂ tensions (c.5%), which are normal for articular cartilage but hypoxic for other cell types, promote a chondrocyte phenotype [21-23]. In addition, however, a pathological role for O₂ has also received considerable attention. Thus abnormally high or low O₂ levels with concomitant alterations in levels of ROS, may be important in disease states such as osteoarthritis [24-26]. O₂ also affects acid-base balance in articular chondrocytes [7,8]. The present findings extend the action of O₂ to include modulation of an additional important ion, ie Ca²⁺, with low O₂ causing intracellular [Ca²⁺] to rise. The O₂ tension at which perturbation of Ca²⁺ requires further definition, it being particularly important to study the likely physiological levels of between 10% and 1%.

Calcium and chondrocyte function

Intracellular Ca²⁺ in chondrocytes, as in other cell types, also has numerous physiological and probably pathological roles [27]. Of particular relevance to chondrocytes is the observation that perturbation of normal Ca²⁺ levels

reduces matrix synthesis [10]. It also affects both chondrocyte differentiation [28] and ageing [11]. Ca^{2+} signalling has been implicated in a range of other chondrocyte functions including mechanotransduction [29-32], volume regulation [33-39] and response to electrical stimulation [40]. It may therefore play a critical role in how joint loading and unloading promotes cartilage health. Intracellular Ca^{2+} elevations, for example, induce chondrogenesis via a calcineurin/NF-AT pathway [41]. Extracellular levels of Ca^{2+} are also important in the longer term, when they too may be involved in alteration of matrix production including proteoglycan synthesis and expression of collagen [42-44] - extracellular Ca^{2+} receptors are present. Ca^{2+} is also implicated in the action of proinflammatory cytokines such as IL-1 and, again therefore, has received attention in the context of joint disease such as osteoarthritis [45].

Crosstalk between oxygen, reactive oxygen species and Ca^{2+}

The elevation of intracellular Ca^{2+} at low O_2 reported here was associated with a fall in ROS and also mitochondrial depolarization. In most cell types, though probably not articular chondrocytes, mitochondria are critical for oxidative phosphorylation and hence central to energy production. They are also involved in Ca^{2+} regulation, acting as a sink of, or sometimes a source for, cytoplasmic Ca^{2+} - Ca^{2+} being released via the mitochondrial permeability transition pore (PTP) [46-48]. ROS are generated during mitochondrial respiration [49,50], as well as at other cellular sites. ROS, of course, can be harmful but have also been implicated in intracellular signalling, regulating redox sensitive enzymes and also ion channels. By these means, ROS may modulate intracellular Ca^{2+} , eg acting via modulation of ryanodine receptors, IP3 receptors, Ca^{2+} pumps and NCE [51-53]. Ca^{2+} uptake by mitochondria may itself alter ROS generation - both reduction of ROS (through dissipation of the negative mitochondrial pd) or their elevation have been reported [54,55]. To a certain extent, the direction of change depends on tissue type and respiratory rate. Another obvious signal is represented by hypoxia-inducible factor (HIF). Stabilization of HIF1 α occurs during hypoxia (eg [6,56,57]) and may affect $[\text{Ca}^{2+}]_i$ through effects calcium channel gene expression and activity [58,59]. There is thus considerable scope for cross-talk between O_2 , ROS and Ca^{2+} , together with the role of mitochondria [51,53,55] but the exact coupling in chondrocytes awaits description.

Reactive oxygen species, mitochondria and regulation of Ca^{2+}

We show here that a fall in ROS during hypoxia correlated with elevation of Ca^{2+} , whilst restoration of ROS

levels to those seen at 20% by three disparate reagents (H_2O_2 , Co^{2+} or antimycin A) all resulted in decreased Ca^{2+} . Hypoxia also induced depolarization of mitochondria, indicative of a reduction in electron flow through the mitochondrial electron transport chain, and hence ROS production. Addition of antimycin A also blocks electron transport to the terminal complexes, acting at the Q_i site of complex III to increase ROS output [8], as also observed in the present work. It is thus likely that reduced production of ROS from mitochondria is involved in the rise in Ca^{2+} , as proposed for O_2 -induced changes in NHE activity and intracellular pH [8]. In the case of H^+ , however, perturbed homeostasis on change in O_2 tension is observed rapidly, within a few minutes [60]. Effects on Ca^{2+} appear to occur over a much longer time course, despite sharing sensitivity to ROS levels. The reason for this is not immediately apparent. It may be that Ca^{2+} homeostasis, as a more critical modulator of chondrocyte function, is better protected than pH. Alternatively, it may be that the mechanism involves genomic effects, such as though involving HIF. In addition, a link between Ca^{2+} and pH in chondrocytes has been shown previously, with alkalinisation causing a rise in Ca^{2+} [61]. Since chondrocytes acidify in response to low O_2 , however, rather than increasing their pH, the hypoxia-induced rise in Ca^{2+} cannot be secondary to changes in pH.

Conclusion

O_2 tension exerts a significant effect on cytoplasmic Ca^{2+} levels of articular chondrocytes, with the proposed mechanism involving ROS from mitochondria. Results emphasise the importance of O_2 to chondrocyte function and that of using realistic O_2 tensions when studying the pathophysiology of articular cartilage.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

RW helped plan the experiments, carried them, analysed the data and helped write the manuscript; JSG planned the experiments, analysed data and prepared the manuscript.

All authors have read and approved the final manuscript.

Acknowledgements

This work was supported by the BBSRC, UK.

Author Details

Department of Veterinary Medicine, University of Cambridge, Madingley Road, Cambridge, CB3 0ES, UK

Received: 21 September 2009 Accepted: 26 April 2010

Published: 26 April 2010

References

1. Silver IA: Measurement of pH and ionic composition of pericellular sites. *Phil Trans Roy Soc B* 1975, **271**:261-272.

2. Zhou S, Chiu Z, Urban JPG: **Factors affecting the oxygen concentration gradient from the synovial surface of articular cartilage to the cartilage-bone interface: a modelling study.** *Arthritis Rheum* 2004, **50**:3915-3924.
3. Lee RB, Urban JPG: **Evidence for a negative Pasteur effect in articular cartilage.** *Biochem J* 1997, **321**:95-102.
4. Grimshaw MJ, Mason RM: **Bovine articular chondrocyte function *in vitro* depends upon oxygen tension.** *Osteoarthritis Cartil* 2000, **8**:386-392.
5. Gibson JS, Milner PI, White R, Fairfax TPA, Wilkins RJ: **Oxygen and reactive oxygen species in articular cartilage: modulators of ionic homeostasis.** *Pflug Archiv* 2008, **455**:563-573.
6. Chandel NS, Schumacker PT: **Cellular oxygen sensing by mitochondria: old questions, new insight.** *J Appl Physiol* 2000, **88**:1880-1889.
7. Milner PI, Fairfax TPA, Browning JA, Wilkins RJ, Gibson JS: **The effect of O₂ tension on pH homeostasis in equine articular chondrocytes.** *Arthritis Rheum* 2006, **54**:3523-3532.
8. Milner PI, Wilkins RJ, Gibson JS: **The role of mitochondrial reactive oxygen species in pH regulation in articular chondrocytes.** *Osteoarthritis Cart* 2007, **15**:735-742.
9. Tattersall AL, Meredith D, Furla P, Shen M-R, Ellory JC, Wilkins RJ: **Molecular and functional identification of the Na⁺/H⁺ exchange isoforms NHE1 and NHE3 in isolated bovine articular chondrocytes.** *Cell Physiol Biochem* 2003, **13**:215-222.
10. Wilkins RJ, Browning JA, Ellory JC: **Surviving in a matrix: membrane transport in articular chondrocytes.** *J Membrane Biol* 2000, **177**:95-108.
11. Nevo Z, Beit-Or A, Eilam Y: **Slowing down aging of cultured embryonal chick chondrocytes by maintenance under lowered oxygen tension.** *Mech Ageing Develop* 1988, **45**:157-165.
12. Sanchez JC, Wilkins RJ: **Mechanisms involved in the increase in intracellular calcium following hypotonic shock in bovine articular chondrocytes.** *Gen Physiol Biophys* 2003, **22**:487-500.
13. Brighton CT, Kitajima T, Hunt RM: **Zonal analysis of cytoplasmic components of articular cartilage chondrocytes.** *Arthritis Rheum* 1984, **27**:1290-1299.
14. Stockwell RA: **Morphometry of cytoplasmic components of mammalian articular chondrocytes and corneal keratocytes: species and zonal variations of mitochondria in relation to nutrition.** *J Anat* 1991, **175**:251-261.
15. Martin JA, Buckwalter JA: **The role of chondrocyte senescence in the pathogenesis of osteoarthritis and in limiting cartilage repair.** *J Bone Joint Surg* 2003, **85A**:106-110.
16. Mignotte F, Champagne A-M, Froger-Gaillard B, Benel L, Gueride M, Adolphe M, Mounolou JC: **Mitochondrial biogenesis in rabbit articular chondrocytes transferred to culture.** *Biol Cell* 1991, **71**:67-72.
17. Etherington PJ, Winlove P, Taylor P, Paleolog E, Miotla JM: **VEGF release is associated with reduced oxygen tensions in experimental inflammatory arthritis.** *Clin Exp Rheumatol* 2002, **20**:799-805.
18. Cernanec J, Guilak F, Weinberg JB, Pisetsky DS, Fermor B: **Influence of hypoxia and reoxygenation on cytokine-induced production of proinflammatory mediators in articular cartilage.** *Arthritis Rheum* 2002, **46**:968-975.
19. Murphy CL, Polak JM: **Control of human articular chondrocyte differentiation by reduced oxygen tension.** *J Cell Physiol* 2004, **199**:451-459.
20. Marcus RE, Srivastava VML: **Effect of low oxygen tension on glucose-metabolising enzymes in cultured articular chondrocytes.** *Proc Soc Exp Biol Med* 1973, **143**:488-491.
21. Murphy CL, Sambanis A: **Effect of oxygen tension on chondrocyte extracellular matrix accumulation.** *Conn Tissue Res* 2001, **42**:87-96.
22. Wang DW, Fermor B, Gimble JM, Awad HA, Guilak F: **Influence of oxygen on the proliferation and metabolism of adipose-derived adult stem cells.** *J Cell Physiol* 2005, **204**:184-191.
23. Betre H, Ong SR, Guilak F, Chilkoti A, Fermor B, Setton LA: **Chondrocyte differentiation of human adipose-derived adult stem cells in elastin-like polypeptide.** *Biomaterials* 2006, **27**:91-99.
24. Henroitin YE, Bruckner P, Pujol JP: **The role of reactive oxygen species in homeostasis and degradation of cartilage.** *Osteoarthritis Cart* 2003, **11**:747-755.
25. Hitchon CA, El-Gabalawy HS: **Oxidation in rheumatoid arthritis.** *Arthritis Res Ther* 2004, **6**:265-278.
26. Henroitin YE, Kurz B, Aigner T: **Oxygen and reactive oxygen species in cartilage degradation: friends or foes?** *Osteoarthritis Cart* 2005, **13**:643-654.
27. Berridge MJ, Bootman MD, Lipp P: **Calcium - a life and death signal.** *Nature* 1998, **395**:645-648.
28. Matta C, Fodor J, Szijgyarto Z, Juhász T, Gergely P, Csernoch L, Zákány R: **Cytosolic free Ca²⁺ concentration exhibits a characteristic temporal pattern during *in vitro* cartilage differentiation: a possible role of calcineurin in Ca-signalling of chondrogenic cells.** *Cell Calcium* 2008, **44**:310-323.
29. Yellowley CE, Jacobs CR, Li Z, Zhou Z, J DH: **Effects of fluid flow on intracellular calcium in bovine articular chondrocytes.** *Am J Physiol* 1997, **273**:C30-C36.
30. Guilak F, Zell RA, Erickson GR, Grande DA, Rubin CT, McLeod KJ, Donahue HJ: **Mechanically induced calcium waves in articular chondrocytes are inhibited by gadolinium and amiloride.** *J Orthop Res* 1999, **17**:421-429.
31. Ushida T, Murata T, Mizuno S, Tateishi T: **Transients of intracellular calcium ion concentrations of cultured bovine chondrocytes loaded with intermittent hydrostatic pressure.** *Mol Cell Biol* 1997, **8**:S405.
32. Browning JA, Saunders K, Urban JPG, Wilkins RJ: **The influence and interactions of hydrostatic pressure and osmotic pressure on the intracellular milieu of chondrocytes.** *Biorheology* 2004, **41**:299-308.
33. Erickson GR, Alexopoulos LG, Guilak F: **Hyper-osmotic stress induces volume change and calcium transients in chondrocytes by transmembrane, phospholipid, and G-protein pathways.** *J Biomech* 2001, **34**:1527-1535.
34. Erickson GR, Northrup DL, Guilak F: **Hypo-osmotic stress induces calcium-dependent actin reorganisation in articular chondrocytes.** *Osteoarthritis Cart* 2003, **11**:187-197.
35. Yellowley CE, Hancox JC, Donahue HJ: **Cell swelling activation of membrane currents, Ca²⁺ transients and regulatory volume decrease in bovine articular chondrocytes.** *J Bone Mineral Res* 2001, **16**:S254.
36. Wilkins RJ, Davies ME, Muzyamba MC, Gibson JS: **Homeostasis of intracellular Ca²⁺ in equine chondrocytes: response to hypotonic shock.** *Equine Vet J* 2003, **35**:439-443.
37. Kerrigan MJP, Hall AC: **The role of [Ca²⁺]_i in mediating regulatory volume decrease in isolated bovine articular chondrocytes.** *J Physiol* 2000, **527P**:42P.
38. Kerrigan MJP, Hall AC: **Control of chondrocyte regulatory volume decrease (RVD) by [Ca²⁺]_i and cell shape.** *Osteoarthritis Cart* 2008, **16**:312-322.
39. Phan MN, Leddy HA, Votta BJ, Kumar S, Levy DS, Lipshutz DB, Lee SH, Liedtke W, Guilak F: **Functional characterisation of TRPV4 as an osmotically sensitive ion channel in porcine articular chondrocytes.** *Arthritis Rheum* 2009, **60**:3028-3037.
40. Xu J, Wang W, Clark CC, Brighton CT: **Signal transduction in electrically stimulated articular chondrocytes involves translocation of extracellular calcium through voltage-gated channels.** *Osteoarthritis Cart* 2009, **17**:397-405.
41. Tomita M, Reinhold MI, Molkenin JD, Naski MC: **Calcineurin and NFAT4 induce chondrogenesis.** *J Biol Chem* 2002, **277**:42214-42218.
42. Shulman HJ, Opler A: **The stimulatory effect of calcium on the synthesis of cartilage proteoglycan.** *Biochem Biophys Res Comm* 1974, **59**:914-919.
43. Benya PD, Shaffer JD: **Dedifferentiated chondrocytes reexpress the differentiated collagen phenotype when cultured in agarose gels.** *Cell* 1982, **30**:215-224.
44. Urban JP, Hall AC, Gehl KA: **Regulation of matrix synthesis rates by the ionic and osmotic environment of articular chondrocytes.** *J Cell Physiol* 1993, **154**:262-270.
45. Pritchard S, Guilak F: **Effects of interleukin-1 on calcium signalling and the increase of filamentous actin in isolated and in situ chondrocytes.** *Arthritis Rheum* 2006, **54**:2164-2174.
46. Herrington J, Park YB, Babcock DF, Hille B: **Dominant role of mitochondria in clearance of large Ca²⁺ loads from rat adrenal chromaffin cells.** *Neuron* 1996, **16**:219-228.
47. Eager KR, Roden LD, Dulhunty AF: **Actions of sulfhydryl reagents on single ryanodine receptor Ca²⁺-release channels from sheep myocardium.** *Am J Physiol* 1997, **272**:C1908-C1918.
48. Boitier E, Rea R, Duchon MR: **Mitochondria exert a negative feedback on the propagation of intracellular Ca²⁺ waves in rat cortical astrocytes.** *J Cell Biol* 1999, **145**:795-808.

49. Kowaltowski AJ, Naia-da-Silva ES, Castilho RF, Vercesi AE: **Ca²⁺-stimulated mitochondrial reactive oxygen species generation and permeability transition are inhibited by dibucaine or Mg²⁺.** *Arch Biochem Biophys* 1998, **359**:77-81.
50. Maciel EN, Vercesi AE, Castilho RF: **Oxidative stress in Ca²⁺-induced membrane permeability transition in brain mitochondria.** *J Neurochemistry* 2001, **79**:1237-1245.
51. Yan Y, Wei CL, Zhang WR, Cheng HP, Liu J: **Cross-talk between calcium and reactive oxygen species signaling.** *Acta Pharmacol Sin* 2006, **27**:821-826.
52. Zima AV, Blatter LA: **Redox regulation of cardiac calcium channels and transporters.** *Cardiovascular Res* 2006, **71**:310-321.
53. Camello-Almarez C, Gomez-Pinilla PJ, Pozo MJ, Camello PJ: **Mitochondrial reactive oxygen species and Ca²⁺ signaling.** *Am J Cell Physiol* 2006, **291**:C1082-C1088.
54. Starkov AA, Chinopoulos C, Fiskum G: **Mitochondrial calcium and oxidative stress as mediators of ischemic brain injury.** *Cell Calcium* 2004, **36**:257-264.
55. Feissner RF, Skalska J, Gaum WE, Sheu S-S: **Crosstalk signaling between mitochondrial Ca²⁺ and ROS.** *Frontiers Bioscience* 2009, **14**:1197-1218.
56. Wang GL, Semenza GL: **Oxygen sensing and response to hypoxia by mammalian cells.** *Redox Report* 1996, **2**:89-96.
57. Fahling M: **Cellular oxygen sensing, signalling and how to survive translational arrest in hypoxia.** *Acta Physiol* 2009, **195**:205-230.
58. Del Toro R, Levitsky KL, Lopez-Barneo J, Chiara MD: **Induction of T-type calcium channel gene expression by chronic hypoxia.** *J Biol Chem* 2003, **278**:22316-22324.
59. Wang J, Weigand L, Lu W, Sylvester JT, Semenza GL, Shimoda LA: **Hypoxia inducible factor 1 mediates hypoxia-induced TRPC expression and elevated intracellular Ca²⁺ in pulmonary arterial smooth muscle cells.** *Circulation Res* 2006, **98**:1528-1537.
60. Gibson JS, McCartney D, Sumpster J, Fairfax TP, Milner PI, Edwards HL, Wilkins RJ: **Rapid effects of hypoxia on H⁺ homeostasis in articular chondrocytes.** *Pflug Archiv* 2009, **458**:1085-1092.
61. Browning JA, Wilkins RJ: **The effect of intracellular alkalinisation on intracellular Ca²⁺ homeostasis in a human chondrocyte cell line.** *Eur J Physiol* 2002, **444**:744-751.

doi: 10.1186/1749-799X-5-27

Cite this article as: White and Gibson, The effect of oxygen tension on calcium homeostasis in bovine articular chondrocytes *Journal of Orthopaedic Surgery and Research* 2010, **5**:27

**Submit your next manuscript to BioMed Central
and take full advantage of:**

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

